

Directions for use



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This following protocol is an example for use in adherent mammalian cells. The cell culture medium in this example uses bicarbonate as a pH-buffer. Cells are cultured in a flask with a standard phenolic style screw cap (i.e. without a membrane) which can be closed air-tight*. This enables 5% CO₂ enriched air to remain present in the flask during storage, preventing a sharp increase in pH due to a drop in CO₂.

- 1. Clean the outside of the container with your disinfectant of choice.
- 2. Dissolve the lyophilized ROKEPIE-FD01 in 2.0ml solvent or culture medium of your choice.
- 3. Add ROKEPIE®-FD01 stock solution in a 1:25 ratio (confluent monolayer) or 1:50 ratio (semi confluent monolayer) to either fresh or already present cell culture medium.
- 4. Incubate the cells in the supplemented medium for a minimum of two hours at 37°C.
- 5. Close the flask air-tight and place the cells in a cold-room or refrigerator (2-8°C).
- After cold-storage, place cells in a humidified incubator with a slightly opened cap and replace the supplemented cell culture medium once cells have re-attached**. This normally takes about 6 – 24 hours. If cells have not detached during cold-storage, the supplemented cell culture medium can be replaced immediately.

*) If no standard phenolic style caps are available, the filter can be covered by parafilm in order to seal the flask air-tight. Wells or dishes can be covered entirely with parafilm, or all around the edges of the lid.

**) Detachment of cells can occur in some cell lines, but does not necessarily indicate they are apoptotic/necrotic. ROKEPIE@-S01 is not toxic at working concentrations and 24 hour exposure to cells at 37°C should not results in loss of viability.



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